

Asymmetric Synthesis of α,α-Disubstituted α-Amino Acids by Diastereoselective Alkylation of Camphor-Based Tricyclic Iminolactone

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A novel and convenient route for the preparation of chiral tricyclic iminolactones 9 and 10 from camphorquinone has been developed. Alkylation of iminolactones 9 and 10 provided iminolactones 16 and 17 in high yields which were, in turn, alkylated again to afford the α,α -disubstituted products in good yields (70–90%) and excellent diastereoselectivities (>98%). Hydrolysis of the alkylated iminolactones furnished the desired α,α -disubstituted α -amino acids in good yields and high enantiomeric excesses with good recovery yields of the chiral auxiliary 12 and 13. The extremely high endo-face selectivity for alkylation is discussed using semiempirical (MOPAC 93) calculations.

Introduction

Optically active nonproteinogenic amino acids¹ are valuable compounds of high interest not only owing to their remarkable pharmacological and biological activities but also for their role as an investigative topographic probe for bioactive conformations of peptides and the mechanisms of enzyme reactions.² There is a growing demand for optically active, ideally enantiomerically pure, uncommon amino acids resulting from the recent progresses in molecular biology and protein engineering technologies.³ Numerous methods to synthesize these compounds have been reported, such as Schöllkopf's bislactim ether 1,⁴ Williams' diphenyloxazinones 2,⁵ Seebach's oxazolidinones 3,⁶ Oba and Alcaraz's diketopiperazine 4⁷ and 5,⁸ Cativiela's pinanone derivative 6,⁹ as well as Nájera's oxazinone 7,¹⁰ and pyrazinone 8.¹¹ There is a promising trend in the area

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of chiral phase-transfer catalyst (PTC) catalyzed α -alkylation of glycine derivatives.¹² Most of these chiral templates, derived from alanine or acting as alanine equivalents, are very useful in the synthesis of α -methyl α -amino acids.¹³ Unfortunately, the majority of them are not readily amenable to the synthesis of non- α -methyl α , α -disubstituted α -amino acids.

Numerous methods have been developed for the synthesis of α -amino acids, while chiral glycine equivalents occupy an important niche in amino acid synthesis.¹⁴ Recently, we have reported two novel chiral tricyclic iminolactones, (1*R*,2*S*,8*S*)-1,11,11-trimethyl-3-oxa-6-azatricyclo[6.2.1.0^{2.7}]undec-6-en-4-one (**9**)¹⁵ and (1*S*,2*R*,8*R*)-8,11,11-trimethyl-3-oxa-6-azatricyclo[6.2.1.0^{2.7}]undec-6-en-4-one (**10**),¹⁶ prepared from (1*R*)-(+)-camphor as glycine equivalents, which have been successfully applied to the asymmetric synthesis of α -monosubstituted α -amino acids. In this paper, we report a new and shorter synthesis of compounds **9** and **10** as well as their application to the asymmetric synthesis of α -amino acids.



Results and Discussion

A new route for the preparation of chiral tricyclic iminolactones **9** and **10** from camphorquinone is illustrated in Scheme



1. The two carbonyl groups of camphorquinone were reduced with sodium borohydride at ca. 0 °C in a diethyl ether/methanol (v/v = 1:1) mixture to afford 2-*exo*-hydroxyepicamphor (12) and 3-exo-hydroxycamphor (13) in a ratio of 1:1.85 as an inseparable mixture. The mixture of hydroxy ketones 12 and 13 was esterified with N-Cbz-glycine, DMAP, and DCC in dry THF to yield again a 1:1.85 ratio of inseparable mixture of regioisomers 14 and 15 in 98% yield as a viscous oil after column chromatography. The deprotection of the Cbz group by catalytic hydrogenation was carried out in absolute ethanol under a hydrogen atmosphere (1 atm) in the presence of 5% palladium on active carbon as a catalyst for 4 h, and the reaction mixture was then stirred for another 12 h at rt after removal of the hydrogen balloon. Fortunately, two easily separable iminolactones 9 and 10 were obtained after flash chromatography in a ratio of 1:1.59 (**9**: $R_f = 0.14$, **10**: $R_f = 0.38$; EtOAc/hexanes = 1:2).

In accord with our previous results, excellent stereoselectivity has been achieved in the alkylation of iminolactones 9 and 10 when LDA was employed as the base to give the monoalkylated products 16 and 17, respectively, as the sole detectable products in good yields (Scheme 2). The alkylation of compounds 16 and 17 was performed under the same conditions used in the alkylation of iminolactones 9 and 10 except 1.3 equiv of LDA was utilized, and the results are collected in Table 1. Interestingly, the enolates of iminolactones 16 and 17 always attack the electrophiles from the endo face regardless the size of the electrophiles and generate the stereocenter with virtually complete diastereoselectivity. Consequently, the configuration of the α,α -disubstitued α -amino acid can be controlled by employing proper order of alkylations of iminolactones 9 and 10.

Encouraged by the above results, alkylation of mixtures of compounds **16** and **20**, as well as **17** and **21** were carried out using the same reaction conditions as above. As anticipated, excellent results were observed as summarized in Table 2.

The chemical shifts of characteristic protons of compounds **18** and **19** are compiled in Table 3.¹⁷ The C₂-H of compound **18a** appears at a much higher field (δ 2.24) than those of the same proton of compounds **18b** and **18c** (δ 4.27 and 4.25) revealing that it is shielded by the phenyl group of the *endo*-benzyl substituent. The C₂-H of compounds **18e** and **18d** appear

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⁽¹⁷⁾ Like their monosubstituted iminolactones, compounds **18** and **19** exhibit similar distinctive proton NMR absorptions, which are unique to the rigid structure systems; see refs 13 and 14.



TABLE 1. Alkylation of Alkylated Tricylic Iminolactone 16 and 17

entry	substrate	R	E ⁺	product	yield ^a (%)	endo/exo ^b	% de ^c
1	16	CH ₃	C ₆ H ₅ CH ₂ Br	18a	80	>99:1	>98
2	16	$C_6H_5CH_2$	CH ₃ I	18b	82	>99:1	>98
3	16	$C_6H_5CH_2$	CH ₂ =CHCH ₂ Br	18c	75	>99:1	>98
4	16	$C_6H_5CH_2$	C ₆ H ₅ CH ₂ Br	18d	$74 \ (90)^d$		
5	16	$CH_2 = CHCH_2$	C ₆ H ₅ CH ₂ Br	18e	78	>99:1	>98
6	16	$CH_2 = CHCH_2$	CH ₃ (CH ₂) ₃ I	18f	80	>99:1	>98
7	17	CH ₃	C ₆ H ₅ CH ₂ Br	19a	75	>99:1	>98
8	17	$C_6H_5CH_2$	CH ₃ I	19b	88	>99:1	>98
9	17	$C_6H_5CH_2$	$CH_2 = CHCH_2Br$	19c	85	>99:1	>98
10	17	$CH_2 = CHCH_2$	C ₆ H ₅ CH ₂ Br	19d	90	>99:1	>98
11	17	$C_6H_5CH_2$	H^+	19e	71	>99:1	>98
12	17	CH_3	CH ₂ =CHCH ₂ Br	19f	75	>99:1	>98

^{*a*} The reported yields are isolated yields after column separation. ^{*b*} The ratios were estimated by NMR integrations of the crude reaction mixtures on a Varian Mercury-400 NMR spectrometer. ^{*c*} The diastereomeric excesses were determined by HPLC analyses of the crude reaction mixture. ^{*d*} The numbers in parentheses are the yields based on recovered starting material.

TABLE 2. Alkylation of Mixtures of Monosubstituted Iminolactones

		$ \begin{array}{c} & & \\ & & \\ & & \\ & & \\ H \end{array} \begin{array}{c} & & \\ & & \\ H \end{array} \begin{array}{c} & & \\ & & \\ & & \\ H \end{array} \begin{array}{c} & & \\ & & \\ & & \\ & & \\ H \end{array} \begin{array}{c} & & \\ & & $	+	1. LDA (1.3 eq.) 2. THF, HMPA 378°C, E ⁺	$\begin{array}{c} 13 \\ 10 \\ 10 \\ 14 \\ 14 \\ 14 \\ 14 \\ 18 \\ 18 \\ 18 \\ 18$		
		$ \begin{array}{c} $	D_{+} A_{H} A_{H	1. LDA (1.3 eq.) 2. THF, HMPA 378°C, E ⁺	$\begin{array}{c} 13 \\ 10 \\ 9 \\ 9 \\ 14 \\ 14 \\ 19 \\ 19 \\ 19 \\ 14 \\ 19 \\ 14 \\ 19 \\ 14 \\ 19 \\ 14 \\ 19 \\ 14 \\ 10 \\ 10 \\ 10 \\ 10 \\ 10 \\ 10 \\ 10$		
entry	substrate	R	E^+	product	yield ^a (%)	endo/exo ^b	% de ^c
1	16 + 20	CH ₃	C6H5CH2	18a	$80 (88)^d$	>99:1	>98
2	16 + 20	C ₆ H ₅ CH ₂	CH ₃	18b	82	>99:1	>98
3	16 + 20	C ₆ H ₅ CH ₂	$CH_2 = CHCH_2$	18c	81	>99:1	>98
4	16 + 20	CH ₃	$CH_2 = CHCH_2$	18g	93	>99:1	>98
5	17 + 21	CH_3	C ₆ H ₅ CH ₂	19a	91	>99:1	>98
6	17 + 21	C ₆ H ₅ CH ₂	CH ₃	19b	83	>99:1	>98
7	17 + 21	C ₆ H ₅ CH ₂	CH ₂ =CHCH ₂	19c	86	>99:1	>98

^{*a*} The reported yields are isolated yields after column separation. ^{*b*} The ratios were estimated by NMR integrations of the crude reaction mixtures on a Varian Mercury-400 NMR spectrometer. ^{*c*} The diastereomeric excesses were determined by HPLC analyses of the crude reaction mixture. ^{*d*} The number in parentheses is the yield based on recovered starting material.

at a still higher field (δ 2.04 and 1.87) than that of compound **18a**, suggesting that the *exo*-allyl is larger than the corresponding methyl group and the *exo*-benzyl is larger still which forces the *endo*-benzyl group more and more downward. Hence, the phenyl group gets closer and closer to C₂-H resulting in more effective shielding. Similar chemical shift changes are observed for the C_{10α}-H of compounds **18a**-e, further supporting the assigned endo-stereochemistry of products **18a**-e. On the other hand, the C₁₂-CH₃ of compounds **18a**-e show an opposite trend, indicating that the methyl group is shielded by the *exo*-benzyl group more and more effectively from compounds **18b** to **18c** and to **18d**. Exactly the same trends can be found for compounds **19a**-e, suggesting that the electrophiles reacted with the enolate

TABLE 3.Chemical Shifts (ppm) of Characteristic Protons of
Compounds 18 and 19

compd	R	Е	$\delta_{\rm C_2-H}$	$\delta_{C_{10\alpha}-H}$	$\delta_{\mathrm{C}_{12}-\mathrm{CH}_3}$
18a	CH ₃	C ₆ H ₅ CH ₂	2.24	0.68 - 0.62	0.75
18b	C ₆ H ₅ CH ₂	CH ₃	4.27	1.33 - 1.29	0.21
18c	C ₆ H ₅ CH ₂	$CH_2 = CHCH_2$	4.25	1.30 - 1.21	0.05
18d	C ₆ H ₅ CH ₂	C ₆ H ₅ CH ₂	1.87	0.48 - 0.42	-0.18
18e	$CH_2 = CHCH_2$	C ₆ H ₅ CH ₂	2.04	0.57 - 0.50	0.73
19a	CH ₃	C ₆ H ₅ CH ₂	2.40	0.66 - 0.62	0.69
19b	C ₆ H ₅ CH ₂	CH ₃	4.43	1.32 - 1.26	0.09
19c	C ₆ H ₅ CH ₂	$CH_2 = CHCH_2$	4.41	1.25 - 1.19	-0.07
19d	$CH_2 = CHCH_2$	C ₆ H ₅ CH ₂	2.18	0.59 - 0.56	0.67
19e	C ₆ H ₅ CH ₂	Н	4.43	1.35 - 1.28	0.71

from the bottom face to afford the endo isomers as the predominant product.



FIGURE 1. X-ray structure of compound 18b.



FIGURE 2. X-ray structure of compound 18d.



FIGURE 3. X-ray structure of compound 18g.

Recrystallization from ethyl acetate and hexanes afforded compounds **18b**, **18d**, **18g**, and **19e**. Their X-ray crystallographic structures are shown in the Figures 1–4, respectively.¹⁸ It is evident that the iminolactone ring, fused with the rigid camphor frame, is in a boat conformation with the C₂-proton and the C_{5endo}-group at the flagpole positions. The X-ray structure of compound **18b** clearly demonstrates that its C₁₂-CH₃ is situated in front of the shielding zone of the phenyl ring of the *exo*-benzyl group, explaining its unusual upfield position in ¹H NMR. The phenyl ring of the *exo*-benzyl group, which



FIGURE 4. X-ray structure of compound 19e.

TABLE 4. Hydrolysis of the Dialkylated Iminolactones 18 and 19

entry	R	E	amino acid (%)	recovered auxiliary (%)	ee ^{<i>a</i>} (%)	$[\alpha]^{22}D^b$
18a	CH ₃	C ₆ H ₅ CH ₂	86	80	98.2	+21.6
18b	C ₆ H ₅ CH ₂	CH ₃	88	82	97.7	-21.5
18c	C ₆ H ₅ CH ₂	$CH_2 = CHCH_2$	82	75	98.1	+26.8
19a	CH ₃	C ₆ H ₅ CH ₂	89	80	98.2	-21.6
19b	C ₆ H ₅ CH ₂	CH_3	89	85	97.7	-21.5
19c	C ₆ H ₅ CH ₂	$CH_2 = CHCH_2$	87	84	98.9	-27.0
19f	CH ₃	$CH_2 = CHCH_2$	90	85	98.2	-28.0

 a ee (%) values are determined by comparison with [α] values in the literature; see the Experimental Section. b The optical rotations were measured in H₂O solution on a Perkin-Elmer PE-241 polarimeter.

shields the C₁₂-CH₃ more effectively, and thus, the signal of this methyl appeared at δ -0.18. Moreover, the C₂-H falls in the shielding cone of the phenyl ring of the *endo*-benzyl group of compound **18d**, making it to appear at δ 1.87, which is considerably higher field than the corresponding C₂-H of other alkylated iminolactones. Consequently, the absolute stereochemistry established by the X-ray structures is in accord with the observed characteristic ¹H NMR absorptions (Table 3). As a result, the attack of the enolate takes place from the endo face is established unequivocally.

Representative dialkylated tricyclic iminolactones **18** and **19** were hydrolyzed with 2 N NaOH at rt for 2 h and then with 6 N HCl at 92 °C for 3 h¹⁰ to afford the corresponding optically active α, α -disubstituented α -amino acids in good yields (Table 4) and enantiomeric excesses with good recovery yields of the chiral auxiliaries **12** and **13**. The configuration of the α -amino acids is in agreement with that assigned to the respective dialkylated tricyclic iminolactone precursors.

Theoretical Investigation

To gain an insight on the mechanism of and to look for an explanation for the high stereoselectivity of the substitution reaction from an energetic point of view, we turned to a systematic quantum chemistry study.¹⁹

⁽¹⁸⁾ The X-ray crystallographic structures of compounds **18b**, **18d**, **18g**, and **19e** have been submitted to the Cambridge Crystallographic Data Centre. The CCDC numbers for compounds **18b**, **18d**, **18g**, and **19e** are CCDC 277545, CCDC 277543, CCDC 277544 and, CCDC 277542, respectively.

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TABLE 5. Energies for Endo and Exo $S_N 2$ Mechanisms $(kcal/mol)^a$

		endo TS			exo TS			
substituent (RX)	$E_{\rm a}$	ΔE	$\Delta E'$	$E_{\rm a}$	ΔE	$\Delta E'$	$\Delta E_{\rm a}$	
CH ₃ I	29.84	26.06	-1.37	34.52	29.92	-0.65	4.68	
CH ₃ CH ₂ I	35.87	32.22	-0.72	41.64	36.24	0.37	5.77	
CH ₃ CH ₂ CH ₂ I	36.12	33.08	0.97	40.79	37.08	0.99	4.67	
CH ₃ CH ₂ CH ₂ CH ₂ I	36.04	32.99	1.71	40.23	36.98	2.11	4.19	
CH ₂ =CHCH ₂ Br	28.83	25.68	-5.69	32.99	29.15	-4.18	4.16	
C ₆ H ₅ CH ₂ Br	29.73	26.48	-5.66	33.75	29.91	-5.70	4.02	

 ${}^{a}E_{a} = E(TS) - E(reaction intermediate); \Delta E = E_{ts} - E_{total}(reactants^*).$ Reactants*: enolate of iminolactone; $\Delta E' = E_{total}(products) - E_{total}(reactants).$ $\Delta E_{a} = E_{a}(exo TS) - E_{a}(endo TS)$

Because of the large amount to be computed, semiempirical method AM1,²⁰ proven to be reliable for the calculation of transition state,²¹ was employed. Full geometry optimization were carried out at the RHF level, using the Broyden–Fletcher–Goldfarb–Shanno function minimizer (BFGS).²² All calculations were performed on an SGI workstation using AM1 computation implemented in MOPAC 93.²³

Monosubstitution. The $S_N 2$ mechanism of monosubstitution of the tricyclic iminolactone was studied first (Scheme 3), and the calculated results are summarized in Table 5.

All the results were obtained in the gas phase. The differences in heat of formation for the reactants and the products are small (Table 5); however, when the difference between the enthalpies of formation for HI (g) and I^- (aq) (about 20 kcal/mol) is taken into account, the reaction become exothermal and highly energetically favored.

The activation energies, calculated without the consideration of solvation effect, for all reactions are found in the range from 28 to 41 kcal/mol. The differences in E_a range from 4.02 to

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5.77 kcal/mol depending on the reaction. This result is in agreement with experimental data in which the endo diastereomer is the predominant product.^{15,16} Since the difference between the enthalpy of formation of the endo product and exo product is small, indicating that the high facial selectivity observed is not controlled by thermodynamics; instead, the S_N2 reaction is most likely kinetically controlled.

The geometry of the intermediate is depicted in Figure 5 for the illustrated reaction (MeI). The intermolecular distances in the intermediate indicate that the approach from the axial (endo) direction is easier than that from the equatorial (exo) direction (3.01 Å against 3.30 Å). This difference in distance decreases in the corresponding transition states (Figure 6). The steric hindrance resulting from the C₁₂-methyl group is presumably the major factor for the observed high facial selectivity. The electrostatic potential isodensity surfaces of the two transition states obtained from ab initio calculations are shown in Figure 7. It is observed that the presence of the methyl group pushes the nucleophilic attack to deviate from the principal axis.

Disubstitution. The substitution reactions of the two isomeric iminolactones were studied next:

Compound 16



Compound 17



The reactions, like in the case of monosubstitution, can proceed easily since the energy barriers are relatively low. Furthermore, in both compounds **16** and **17**, the *endo* attack is favored by kinetics. Figures 8 and 9 show, respectively, two examples of the transition state for reactions of compounds **16** and **17**. Compared with monosubstitution, the distances between the leaving atom (X) and its binding carbon, and between this carbon atom and the enolate, are both slightly elongated, which might be due to the presence of the first substituent. In Table 6, the energy differences between these two nucleophilic attacks for compounds **16** and **17** are compared. The differences range from 1.32 to 5.57 kcal/mol depending on the reactions, which

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FIGURE 5. Geometries of endo and exo intermediates: (left) axial nucleophilic attack; (right) equatorial nucleophilic attack.



FIGURE 6. Geometries of endo and exo transition states: (left) axial nucleophilic attack; (right) equatorial nucleophilic attack.



FIGURE 7. Electrostatic potential isodensity surfaces of two transition states (equatorial and axial nucleophilic attacks), obtained from ab initio calculations.

are similar to the situations for monosubstitution reactions. The results suggest that the attack is more favored from the endo direction to furnish the major product with R' at the axial position. This is in good agreement with experimental results and can explain the high stereoselectivity of this type of reaction.

Ab Initio Calculations. To validate the AM1 semiempirical results, ab initio calculations for two transition states were carried out at the HF level using the DFT method. The B3LYP (Beck's three-parameter hybrid exchange function and the Lee– Yang–Parr correlation function) and the Pople 6-31G(d) basis set were used for the calculations except for the I atom, which

was treated with the 3-21G* basis set. All calculations were performed with Gaussian 98 software at an SGI Octane 2 workstation. The geometries obtained from this computation, shown in Figure 10, are very close to those optimized (Figure 6, 2.43 Å) using the AM1 method.

The structures of both transition states have been verified by the calculation of frequency. For each geometry, we obtained one imaginary frequency (-338.98 cm^{-1} for axial attack TS and -383.37 cm^{-1} for equatorial attack TS). Moreover, the energy difference between the two transition states (axial and equatorial attack) is 3.30 kcal/mol ($\Delta E = E_e - E_a$). In AM1 calculations for this system, the energy difference is 3.86 kcal/



FIGURE 8. Geometries of transition states for alkylation of compound 16 ($R = CH_2CH=CH_2$, R' = n-Bu): (left) axial nucleophile attack; (right) equatorial nucleophile attack.



FIGURE 9. Geometries of transition states for alkylation of isomer 2 ($R = CH_2CH=CH_2$, $R' = CH_2C_6H_5$): (left) axial nucleophilic attack; (right) equatorial nucleophilic attack.

TABLE 6.	Energy	Difference	between	Exo and	Endo	Attack	(kcal/mol)
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reaction compd 16					reaction c	ompd 17			
R	R'	$\Delta E_{ m exo}$	$\Delta E_{ m endo}$	$\Delta(\Delta E)^a$	R	R′	$\Delta E_{ m exo}$	$\Delta E_{ m endo}$	$\Delta(\Delta E)^a$
CH ₃	CH ₂ C ₆ H ₅	35.58	32.25	3.33	CH ₃	CH ₂ C ₆ H ₅	35.40	32.15	3.25
CH ₂ =CHCH ₂	CH ₂ C ₆ H ₅	38.80	35.91	2.89	$CH_2 = CHCH_2$	CH ₂ C ₆ H ₅	38.25	36.10	2.15
CH ₂ =CHCH ₂	<i>n</i> -Bu	41.19	39.62	1.57	CH ₂ C ₆ H ₅	CH ₂ C ₆ H ₅	36.77	33.05	3.72
CH ₂ C ₆ H ₅	CH ₃	36.63	31.06	5.57	CH ₂ C ₆ H ₅	$CH_2 = CHCH_2$	37.42	33.76	3.66
CH ₂ C ₆ H ₅	CH ₂ =CHCH ₂	37.31	35.99	1.32					
CH ₂ C ₆ H ₅	CH ₂ C ₆ H ₅	37.02	34.77	2.25					

 $^{a}\Delta(\Delta E) = \Delta E_{\rm exo} - \Delta E_{\rm endo}.$



FIGURE 10. Geometries of transition states obtained from ab initio calculations.

mol. Therefore, the results obtained from AM1 computations can be considered as valid. Furthermore, the systematic errors should be reduced again for a such comparative study.

Conclusion

In conclusion, we have developed an efficient and practical method for the preparation of α , α -disubstituented α -amino acids

starting from inexpensive and readily available (1R)-(+)camphor. Good chemical yields and excellent diastereoselectivity were realized to produce the α,α -disubstituted α -amino acids in high optical purity. The fact that the chiral auxiliaries can be recycled without loss of optical integrity renders the present method an economical method for the preparation of α,α -disubstituted α -amino acids. Consequently, our method makes it possible to synthesize a wide range of α,α -disubstituted α -amino acids of either stereochemistry.

Experimental Section

General Methods. All alkylation reactions were conducted in flame-dried, long-necked (15 cm), round-bottomed flasks fitted with rubber septa under an argon atmosphere. Solvents and reagents were dried prior to use as required. Diisopropylamine and hexamethylphosphoric triamide (HMPA) were distilled from calcium hydride immediately prior to use, THF was distilled from sodium benzophenone ketyl, and *n*-butylithium in hexanes (nominally 1.6 M) was purchased from a commercial supplier and titrated before each use. Thin-layer chromatography plates were visualized by exposure to ultraviolet light and/or immersion in a staining solution (phosphomolybdic acid) followed by heating on a hot plate. Flash chromatography was carried out utilizing silica gel 60, 70-230 mesh ASTM. Melting points are uncorrected. ¹H NMR spectra were recorded at 400 MHz; ¹³C NMR spectra were recorded at 100 MHz. Chloroform ($\delta = 7.27$ ppm) or deuterium oxide ($\delta = 4.60$ ppm) was used as internal standard in ¹H NMR spectra. The center peak of deuteriochloroform ($\delta = 77.0$ ppm) was used as internal standard in ¹³C NMR spectra. The optical rotations were measured in CHCl₃ solution with a cuvette of 1 dm length. For chiral alkylations, diastereomeric ratios were determined by the integration of the ¹H NMR spectra. The ee value of the α -amino acids obtained from hydrolysis of the alkylated iminolactones was determined by measuring the optical rotation or by HPLC analysis on a Crownpak CR(+) column.

(15,35,4*R*)-3-Hydroxy-4,7,7-trimethylbicyclo[2.2.1]heptan-2one (12) and (1*R*,3*R*,4*S*)-3-Hydroxy-1,7,7-trimethylbicyclo[2.2.1]heptan-2-one (13). To a solution of camphorquinone 11 (4.986 g 30.0 mmol) in a mixture of ether (30 mL) and methanol (30 mL), cooled in an ice bath, was added sodium borohydride (0.298 g, 7.88 mmol, 1.05 equiv) in small batches, and the reaction mixture was stirred in an ice bath for 30 min by which time the color of the reaction changed into pale yellow. The reaction was then washed with cold water (3 × 10 mL), dried (MgSO₄), filtered, and the organic layer was evaporated to give a white powder as an inseparable mixture of two hydroxy ketones. The ¹H NMR of the crude reaction indicated that the hydroxyl ketones were pure enough for the subsequent reactions and therefore was used without further purification (4.39 g, 87%; **12/13** = 1:1.85).

(1R,2S,4S)-Benzyloxycarbonylaminoacetic Acid 1,7,7-Trimethyl-3-oxobicyclo[2.2.1]hept-2-yl Ester (14) and (1S,2R,4R)-Benzyloxycarbonylaminoacetic Acid 4,7,7-Trimethyl-3-oxobicyclo-[2.2.1]hept-2-yl Ester (15). A solution of the mixture of 2-exohydroxyepicamphor (12) and 3-exo-hydroxycamphor (13) (5.046 g, 30.0 mmol), Cbz-glycine (6.897 g, 33.0 mmol, 1.1 equiv), and 4-(N,N-dimethylamino)pyridine (DMAP, 1.833 g, 15.0 mmol, 0.5 equiv) in THF (120 mL) in a 250 mL flask was stirred at 0 °C for 15 min, and a solution of dicyclohexylcarbodiimide (DCC, 9.285 g, 45.0 mmol, 1.5 equiv) in THF (30 mL) was then added dropwise to the reaction via a syringe pump over 40 min. The reaction was stirred at 0 °C for 2 h and then at rt for 16 h. Precipitated 1,3dicyclohexylurea was removed by filtration and the filtrate concentrated under reduce pressure. Purification of the residue by flash column chromatography (EtOAc/hexanes = 1:8) gave an inseparable mixture of two esters as a colorless viscous oil (10.56 g, 98%; 14/15 = 1:1.85).

(1*R*,2*S*,8*S*)-1,11,11-Trimethyl-3-oxa-6-azatricyclo[6.2.1.0^{2,7}]undec-6-en-4-one (9) and (1*S*,2*R*,8*R*)-8,11,11-Trimethyl-3-oxa**6-azatricyclo[6.2.1.0**^{2,7}**]undec-6-en-4-one (10).** A 100 mL twonecked flask containing the mixture of esters **14** and **15** (5.408 g, 15 mmol) and 5% palladium on activated carbon (0.604 g) was evacuated and filled with hydrogen three times. Freshly dried ethanol (40 mL) was added to the mixture followed by evacuation and filling with hydrogen one more time. The mixture was stirred under hydrogen atmosphere (1 atm) at rt (ca. 25 °C) for 14 h. The catalyst was removed by filtration, and the filtrate was concentrated to afford the crude products. The residue was purified by column chromatography (EtOAc/hexanes = 1:4) to furnish the desired iminolactones **9** (0.78 g, 25%) and **10** (1.24 g, 40%).

Iminolactone **9**: mp 63–64 °C; $[\alpha]^{22}_{D}$ –265.6 (c = 2.34, CHCl₃); ¹H NMR δ 4.52 (d, J = 18 Hz, 1H), 4.32 (d, J = 1.6 Hz), 3.90 (dd, J = 1.6, 18 Hz, 1H), 2.45 (d, J = 4.4 Hz, 1H), 2.05–1.98 (m, 1H), 1.95–1.88 (m, 1H), 1.59–1.52 (m, 1H), 1.43–1.36 (m, 1H), 1.09 (s, 3H), 0.98 (s, 3H), 0.86 (s, 3H); ¹³C NMR δ 181.8, 168.8, 81.7, 53.2, 52.5, 49.4, 48.9, 34.0, 21.6, 20.0, 19.3, 9.8; IR (NaCl, CHCl₃) 2962 (m), 1751 (s), 1695 (m) cm⁻¹; HRMS *m/e* calcd for C₁₂H₁₇NO₂ M⁺ 207.1268, found M⁺ 207.1264. Anal. Calcd for C₁₂H₁₇NO₂: C, 69.48; H, 8.27; N, 6.75. Found: C, 69.44; H, 8.25; N, 6.72.

Iminolactone **10**: mp 103–104 °C; $[\alpha]^{22}_{D}$ +266.3 (c = 1.35, CHCl₃); IR (NaCl, CHCl₃): 2972 (m), 1758 (s), 1685 (m) cm⁻¹; ¹H NMR δ 4.56 (d, J = 18 Hz, 1H), 4.49 (d, J = 1.6 Hz, 1H), 3.95–3.90 (dd, J = 18 Hz, 1.6 Hz), 2.28 (d, J = 4.8 Hz), 2.13–2.06 (m, 1H), 1.83–1.75 (m, 1H), 1.62–1.55 (m, 1H), 1.41–1.34 (m, 1H), 1.07 (s, 3H), 0.98 (s, 3H), 0.81 (s, 3H); ¹³C NMR δ 184.0, 169.2, 79.9, 52.9, 49.3, 47.7, 29.7, 25.6, 20.2, 19.9, 10.1; MS m/z 207 (M⁺, 64.0), 192 (5.1), 179 (100.0), 164 (13.7), 150 (32.0), 136 (31.9), 111 (51.2), 110 (22.7), 82 (44.0), 69 (54.7), 55 (15.4), 53 (10.7); HRMS m/e calcd for C₁₂H₁₇NO₂ M⁺ 207.1268, found M⁺ 207.1259. Anal. Calcd for C₁₂H₁₇NO₂: C, 69.48; H, 8.27; N, 6.75. Found: C, 69.54; H, 8.26; N, 6.44.

General Procedure for the Alkylation of Tricyclic Iminolactones (9, 10, 16, and 17). Diisopropylamine (216 μ L, 1.54 mmol, 1.1 equiv) was added to a solution of dry THF (1.2 mL) and *n*-BuLi (1.6 *M*, 962 μ L, 1.54 mmol, 1.1 equiv) at -30 °C and was then stirred at -30 °C for 30 min under an argon atmosphere.

Iminolactone (292 mg, 1.4 mmol) in dry THF (10 mL) was added dropwise over a period of 10 min to the above freshly prepared LDA solution via a syringe at -30 °C, and the resulting solution was stirred at -30 °C for 90 min. After the addition of HMPA (0.73 mL, 4.2 mmol, 3 equiv), the reaction mixture was cooled to -78 °C. A 0 °C solution of alkyl halide (4.2 mmol, 3 equiv) in dry THF (10 mL) was added to the reaction mixture via a syringe pump with the needle contacting the wall of the neck of the reaction vessel over 10 min. The well-stirred reaction was kept at -78 °C for 12 h.

Aqueous acetic acid solution (2 M, 2 mL) was added to the mixture to quench the reaction. The reaction was warmed to room temperature, washed with satd aqueous LiCl (3×3 mL), dried (MgSO₄), filtered, and concentrated to give the crude product. The crude reaction was purified by column chromatography to yield the purified products.

(1*R*,2*S*,5*R*,8*S*)-5-Benzyl-1,5,11,11-tetramethyl-3-oxa-6azatricyclo[6.2.1.0^{2,7}]undec-6-en-4-one (18a): 347 mg (yield 80%); [α]²²_D -32.7 (c = 2.29, CHCl₃); R_f 0.33 (hexanes/EtOAc = 2/1); IR (NaCl, CHCl₃) 2963 (s), 2882 (m), 1741 (s), 1698 (m) cm⁻¹; ¹H NMR δ 7.30-7.15 (m, 5H), 3.34 (d, J = 13.6 Hz, 1H), 2.92 (d, J = 13.6 Hz, 1H), 2.32 (d, J = 4.4 Hz, 1H), 2.24 (s, 1H), 1.90-1.78 (m, 1H), 1.72 (s, 3H), 1.68-1.54 (m, 1H), 1.41-1.34 (m, 1H), 0.86 (s, 3H), 0.81 (s, 3H), 0.75 (s, 3H), 0.68-0.62 (m, 1H); ¹³C NMR δ 178.5, 174.1, 136.2, 130.5, 128.6, 127.4, 81.5, 64.9, 54.0, 49.0, 48.0, 46.0, 34.5, 29.5, 21.1, 19.8, 19.2, 9.4; MS m/z311 (M⁺, 1.3), 282 (0.8), 267 (62.0), 252 (30.1), 220 (6.7), 192 (18.5), 176 (100.0), 148 (18.2), 117 (8.6), 91 (41.6), 83 (7.5), 55 (6.9); HRMS m/e calcd for C₂₀H₂₅NO₂: C, 77.14; H, 8.09; N, 4.50. Found: C, 77.10; H, 8.24; N, 4.45. (1*R*,2*S*,5*S*,8*S*)-5-Benzyl-1,5,11,11-tetramethyl-3-oxa-6azatricyclo[6.2.1.0^{2.7}]undec-6-en-4-one (18b): 356 mg (yield 82%); [α]²²_D -70.7 (c = 1.27, CHCl₃); R_f 0.34 (hexanes/EtOAc = 2/1); mp 118–120 °C; IR (NaCl, CHCl₃) 3032 (ms), 2965 (m), 1743 (s), 1701 (m) cm⁻¹; ¹H NMR δ 7.34–7.13 (m, 5H), 4.27 (s, 1H), 3.50–3.47 (d, J = 13.2 Hz, 1H), 3.13–3.09 (d, J = 13.2 Hz, 1H), 2.29 (d, J = 4.4 Hz, 1H), 1.98–1.79 (m, 2H), 1.59 (s, 1H), 1.46 (m, 1H), 1.41–1.34 (s, 3H), 1.33–1.29 (m, 1H), 0.96 (s, 3H), 0.84 (s, 3H), 0.21 (s, 3H); ¹³C NMR δ 176.6, 173.0, 136.9, 131.3, 127.6, 126.4, 80.5, 63.7, 53.8, 49.6, 48.1, 46.7, 34.5, 23.8, 21.2, 19.3, 19.1, 9.7; MS m/z 311 (M⁺, 43.8), 296 (2.2), 267 (47.1), 252 (30.4), 220 (34.6), 192 (22.4), 176 (100.0), 148 (22.3), 117 (11.2), 91 (55.7), 83 (15.8), 55 (12.0); HRMS m/e calcd for C₂₀H₂₅NO₂ M⁺ 311.1886, found M⁺ 311.1896. Anal. Calcd for C₂₀H₂₅NO₂: C, 77.14; H, 8.09; N, 4.50. Found: C, 77.21; H, 8.15; N, 4.60.

(1R,2S,5S,8S)-5-Allyl-5-benzyl-1,11,11-trimethyl-3-oxa-6azatricyclo[6.2.1.0^{2,7}]undec-6-en-4-one (18c): 354 mg (yield 75%); $[\alpha]^{22}_{D}$ -12.0 (c = 1.39, CHCl₃); R_f 0.49 (hexanes/EtOAc = 2/1); ¹H NMR δ 7.29-7.11 (m, 5H), 5.89-5.76 (m, 1H), 5.22-5.17 (m, 2H), 4.25 (s, 1H), 3.48 (d, J = 12.8 Hz, 1H), 3.09 (d, J = 12.8Hz), 2.67 (dd, J = 13.6, 7.2 Hz, 1H), 2.44 (dd, J = 13.6, 7.6 Hz, 1H), 2.27 (d, J = 4.8 Hz, 1H), 1.97–1.89 (m, 1H), 1.83–1.72 (m, 1H), 1.54-1.46 (m, 1H), 1.30-1.21 (m, 1H), 0.90 (s, 3H), 0.80 (s, 3H), 0.05 (s, 3H); ¹³C NMR δ 177.2, 172.1, 136.5, 131.7, 131.3, 127.6, 126.3, 120.1, 81.2, 70.0, 53.8, 49.3, 47.9, 45.7, 43.1, 34.6, 21.5, 19.1, 18.7, 9.4; MS *m*/*z* 337 (M⁺, 22.6), 293 (85.8), 278 (38.7), 252 (26.5), 250 (22.2), 202 (78.3), 174 (12.2), 160 (15.0), 128 (12.6), 91 (100.0), 83 (16.0), 55 (20.4); HRMS m/e calcd for C222H27-NO₂ M⁺ 337.2042, found M⁺ 337.2050. Anal. Calcd for C₂₂H₂₇-NO2: C, 78.30; H, 8.06; N, 4.15. Found: C, 78.26; H, 8.12; N, 4.30.

(1*R*,2*S*,8*S*)-5,5-Dibenzyl-1,11,11-trimethyl-3-oxa-6-azatricyclo-[6.2.1.0^{2,7}]undec-6-en-4-one (18d). 400 mg (yield 74%); $[\alpha]^{22}_{\rm D}$ –41.9 (*c* = 1.03, CHCl₃); mp 146–147 °C; IR (NaCl, CHCl₃) 2960 (ms), 2928 (m), 1738 (s), 1705 (m) cm⁻¹; ¹H NMR δ 7.29–7.26 (m, 6H), 7.20–7.11 (m, 4H), 3.68 (d, *J* = 12.8 Hz, 1H), 3.54 (d, *J* = 12.8 Hz, 1H), 3.18 (d, *J* = 12.8 Hz, 1H), 3.02 (d, *J* = 12.8 Hz, 1H), 2.16 (d, *J* = 4.8 Hz, 1H), 1.87 (s, 1H), 1.77–1.69 (m, 1H), 1.50–1.42 (m, 1H), -0.18 (s, 3H); ¹³C NMR δ 178.5, 173.0, 136.4, 135.9, 131.4, 130.7, 128.6, 127.9, 127.4, 126.6, 81.0, 69.6, 54.0, 48.5, 48.1, 47.8, 46.1, 34.6, 21.1, 19.0, 18.5, 9.2; MS *m*/*z* 387 (M⁺, 14.6), 343 (56.1), 328 (36.4), 296 (12.3), 252 (100.0), 224 (16.1), 186 (5.1), 160 (2.6), 115 (7.2), 91 (64.6), 83 (10.1), 55 (6.7); HRMS *m*/*e* calcd for C₂₆H₂₉NO₂ M⁺ 387.2205, found M⁺ 387.2199.

(1R,2S,5R,8S)-5-Allyl-5-benzyl-1,11,11-trimethyl-3-oxa-6azatricyclo[6.2.1.0^{2,7}]undec-6-en-4-one (18e): 368 mg (yield 78%); $[\alpha]^{22}_{D}$ -15.3 (c = 2.13, CHCl₃); R_f 0.26 (hexanes/EtOAc = 10/1); IR (NaCl, CHCl₃) 3074 (s), 2961 (m), 1741 (s), 1702 (m) cm⁻¹; ¹H NMR δ 7.29-7.27 (m, 3H), 7.15-7.13 (m, 2H), 5.75-5.67 (m, 1H), 5.23-5.06 (m, 2H), 3.81-3.35 (d, J = 13.6 Hz, 1H), 3.05–2.99 (dd, J = 12.8, 7.6 Hz, 1H), 2.96–2.92 (d, J = 13.6 Hz, 1H), 2.74-2.69 (dd, J = 12.8, 6.8 Hz, 1H), 2.32-2.31 (d, J = 4.8Hz, 1H), 2.04 (s, 1H), 1.81–1.78 (m, 1H), 1.60–1.56 (m, 1H), 1.37-1.32 (m, 1H), 0.84 (s, 3H), 0.77 (s, 3H), 0.73 (s, 3H), 0.57-0.50 (m, 1H); ¹³C NMR δ 179.1, 172.8, 135.8, 133.1, 130.4, 128.5, 127.3, 119.3, 81.0, 68.6, 53.9, 48.7, 47.8, 46.7, 45.3, 34.4, 21.0, 20.1, 19.0, 9.2; MS m/z 337 (M⁺, 57.9), 281 (18.2), 266 (22.1), 246 (23.6), 202 (52.5), 160 (10.6), 146 (4.2), 107 (10.8), 83 (100.0), 47 (19.8); HRMS m/e calcd for C₂₂H₂₇NO₂ M⁺ 337.2042, found M⁺ 337.2034.

(1*R*,2*S*,5*S*,8*S*)-5-Allyl-5-butyl-1,11,11-trimethyl-3-oxa-6azatricyclo[6.2.1.0^{2,7}]undec-6-en-4-one (18f): 340 mg (yield 80%); [α]²²_D -12.0 (c = 1.14, CHCl₃); IR (NaCl, CHCl₃) 2959 (ms), 2875 (m), 1741 (s), 1701 (m) cm⁻¹; ¹H NMR δ 5.80-5.73 (m, 1H), 5.21-5.16 (d, J = 17.2 Hz, 1H), 5.09-5.07 (d, J = 10.4 Hz, 1H), 4.36 (s, 1H), 2.88-2.83 (ddd, J = 13.6, 6.8, 0.8 Hz, 1H), 2.66-2.60 (ddd, J = 13.6, 7.2, 0.8 Hz, 1H), 2.41-2.40 (m, 1H), 2.02–1.73 (m, 3H), 1.59–1.51 (m, 2H), 1.41–1.29 (m, 4H), 1.06 (s, 3H), 0.99 (s, 1H), 0.96 (s, 3H), 0.91–0.87 (m, 3H), 0.82 (s, 3H); 13 C NMR δ 177.6, 172.2, 133.6, 118.8, 80.9, 66.5, 53.8, 49.8, 48.1, 42.9, 36.4, 34.6, 26.1, 22.8, 21.3, 20.1, 19.4, 13.8, 9.7; MS *m*/*z* 303 (M⁺, 51.2), 275 (27.2), 247 (100.0), 216 (62.5), 202 (23.8), 190 (22.7), 164 (22.7), 152 (11.1), 107 (16.7), 83 (24.4), 55 (23.2); HRMS *m*/*e* calcd for C₁₉H₂₉NO₂ 303.2198, found 303.2199. Anal. Calcd for C₁₉H₂₉NO₂: C, 75.21; H, 9.63; N, 4.62; Found: C, 75.22; H, 9.74; N, 4.6.

(1*R*,2*S*,5*R*,8*S*)-5-Allyl-1,5,11,11-tetramethyl-3-oxa-6-azatricyclo-[6.2.1.0^{2,7}]undec-6-en-4-one (18g): 340 mg (yield 93%); [α]²²_D –45.1 (c = 0.41, CHCl₃); R_f 0.33 (hexanes/EtOAc = 2/1); mp 76– 77 °C; ¹H NMR NMR δ 5.80–5.71 (m, 1H), 5.18–5.12 (m, 2H), 4.46 (s, 1H), 2.54–2.49 (ddd, J = 14.0, 7.6, 0.8 Hz, H), 2.41– 2.36 (dd, J = 13.6, 7.6 Hz, H), 2.40 (d, J = 4.4, 1H), 2.04–1.82 (m, 2H), 1.62–1.52 (m, 4H), 1.42–1.35 (m, 1H), 1.07 (s, 3H), 0.96 (s, 3H), 0.84 (s, 3H); ¹³C NMR δ 178.2, 172.9, 131.6, 119.8, 81.6, 62.8, 53.8, 49.8, 48.1, 41.9, 34.5, 26.5, 21.4, 19.8, 19.3, 9.7; MS m/z 261 (M⁺, 9.4), 232 (9.3), 217 (90.5), 202 (76.3), 176 (100.0), 174 (41.7), 148 (36.1), 107 (20.0), 83 (41.5), 55 (35.0), 53 (21.7); HRMS m/e calcd for C₁₆H₂₃NO₂ 261.1722, found 261.1728.

(1*S*, 2*R*, 5*S*, 8*R*)-5-Benzyl-5,8,11,11-tetramethyl-3-oxa-6azatricyclo[6.2.1.0^{2.7}]undec-6-en-4-one (19a): 327 mg (yield 75%); [α]²²_D +61.7 (c = 1.16, CHCl₃); R_f 0.56 (hexanes/EtOAc = 2/1); IR (NaCl, CHCl₃) 3061 (s), 3028 (m), 1741 (s), 1695 (m) cm⁻¹; ¹HNMR δ 7.28–7.26 (m, 3H), 7.12–7.09 (m, 2H), 3.34–3.31 (d, J = 13.2 Hz, 1H), 2.96–2.92 (d, J = 13.2 Hz, 1H), 2.40 (s, 1H), 1.81–1.72 (m, 5H), 1.62–1.58 (m, 1H), 1.42–1.34 (m, 1H), 1.04 (s, 3H), 0.84 (s, 3H), 0.69 (s, 3H), 0.66–0.62 (m, 1H); ¹³C NMR δ 179.6, 174.2, 136.2, 130.3, 128.5, 127.2, 79.5, 65.0, 52.5, 48.0, 47.3, 45.8, 29.6, 28.9, 28.8, 19.9, 19.1, 10.2; MS *m/z* 311 (M⁺, 0.8), 283 (2.4), 267 (100.0), 239 (72.3), 224 (18.3), 192 (29.6), 176 (61.4), 148 (48.1), 117 (11.9), 91 (54.8), 83 (93.1), 69 (17.6); HRMS *m/e* calcd for C₂₀H₂₅NO₂ M⁺ 311.1895, found M⁺ 311.1885. Anal. Calcd for C₂₀H₂₅NO₂: C, 77.14; H, 8.09; N, 4.50. Found: C, 77.15; H, 8.14; N, 4.51.

(1*S*, 2*R*, 5*R*, 8*R*)-5-Benzyl-5,8,11,11-tetramethyl-3-oxa-6azatricyclo[6.2.1.0^{2,7}]undec-6-en-4-one (19b). 383 mg (yield 88%); [α]²²_D +48.2 (c = 1.30, CHCl₃), R_f 0.46 (hexanes/EtOAc = 2/1); mp 100-101 °C; ¹H NMR δ 7.36-7.12 (m, 5H), 4.43 (s, 1H), 3.48 (d, J = 12.8 Hz, 1H), 3.13 (d, J = 12.8 Hz, 1H), 2.04 (d, J = 4.8 Hz, 1H), 2.00-1.94 (m, 1H), 1.72-1.65 (m, 1H), 1.56-1.49 (m, 1H), 1.46 (s, 3H), 1.32-1.26 (m, 1H), 0.98 (s, 3H), 0.82 (s, 3H), 0.09 (s, 3H); ¹³C NMR δ 177.8, 173.2, 136.9, 78.5, 63.6, 52.9, 48.2, 47.7, 46.7, 28.9, 25.8, 23.6, 20.0, 18.5, 10.0; MS *m*/z 311 (M⁺, 21.9), 283 (6.9), 267 (100.0), 252 (59.2), 239 (76.2), 224 (17.0), 192 (35.7), 176 (54.9), 148 (41.2), 117 (16.7), 91 (86.3), 77 (11.4), 69 (17.9); HRMS *m/e* calcd for C₂₀H₂₅NO₂ M⁺ 311.1895, found M⁺ 311.1890. Anal. Calcd for C₂₀H₂₅NO₂: C, 77.14; H, 8.09; N, 4.50. Found: C, 77.10; H, 8.24; N, 4.48.

(1*S*,2*R*,5*R*,8*R*)-5-Allyl-5-benzyl-8,11,11-trimethyl-3-oxa-6azatricyclo[6.2.1.0^{2.7}]undec-6-en-4-one (19c): 401 mg (yield 85%); [α]²²_D +32.7 (c = 1.19, CHCl₃); R_f 0.60 (hexanes/EtOAc = 4/1); mp 62-63 °C^{: 1}H NMR δ 7.29-7.27 (m, 2H), 7.14-7.09 (m, 3H), 5.88-5.76 (m, 1H), 5.17-5.05 (m, 2H), 4.41 (s, 1H), 3.48-3.45 (d, J = 12.4 Hz, 1H), 3.11-3.08 (d, J = 12.4 Hz, 1H), 1.96-1.87 (m, 2H), 1.71-1.62 (m, 1H), 1.52-1.44 (m, 1H), 1.25-1.19 (m, 1H), 0.97 (s, 3H), 0.77 (s, 3H), -0.07 (s, 3H); ¹³C NMR δ 178.4, 172.3, 136.6, 131.9, 131.6, 127.5, 126.4, 120.0, 79.4, 67.1, 53.0, 48.1, 47.6, 45.8, 42.9, 29.3, 26.0, 20.0, 18.2, 10.1; MS m/z 337 (M⁺, 18.1), 293 (100.0), 265 (33.7), 246 (18.0), 202 (30.7), 174 (19.9), 150 (12.2), 120 (7.7), 91 (47.7), 69 (19.6), 55 (5.4); HRMS m/e calcd for C₂₂H₂₇NO₂ M⁺ 337.2042, found M⁺ 337.2040. Anal. Calcd for C₂₂H₂₇NO₂: C, 78.30; H, 8.06; N, 4.15. Found: C, 78.26; H, 8.08; N, 4.23.

(1*S*,2*R*,5*S*,8*R*)-5-Allyl-5-benzyl-8,11,11-trimethyl-3-oxa-6azatricyclo[6.2.1.0^{2.7}]undec-6-en-4-one (19d): 425 mg (yield 90%); [α]²²_D +35.5 (c = 1.38, CHCl₃); *R*_f 0.41 (hexanes/EtOAc = 10/1); IR (NaCl, CHCl₃) 2960 (m), 2926 (m), 1741 (s), 1699 (m) cm⁻¹; ¹H NMR δ 7.29–7.25 (m, 3H), 7.13–7.11 (m, 2H), 5.72–5.68 (m, 1H), 5.21–5.16 (m, 1H), 5.07–5.03 (m, 1H), 3.38–3.34 (d, *J* = 13.2 Hz, 1H), 3.04–2.95 (m, 2H), 2.78–2.73 (m, 1H), 2.18 (s, 1H), 1.77–1.70 (m, 2H), 1.61–1.55 (m, 1H), 1.38–1.34 (m, 1H), 1.04 (s, 3H), 0.82 (s, 3H), 0.67 (s, 3H), 0.59–0.56 (m, 1H); ¹³C NMR δ 180.2, 173.0, 135.9, 133.3, 130.3, 128.4, 127.2, 119.1, 79.1, 68.7, 52.7, 47.8, 47.1, 46.7, 45.2, 31.4, 28.8, 25.8, 22.5, 19.9, 19.6, 10.1; MS *m*/*z* 337 (M⁺, 36.7), 293 (50.5), 265 (48.9), 252 (20.1), 202 (49.0), 174 (41.6), 150 (18.5), 120 (14.2), 91 (100.0), 83 (87.9), 69 (37.6); HRMS *m*/*e* calcd for C₂₂H₂₇NO₂ M⁺ 337.2042, found M⁺ 337.2043.

(1*S*,2*R*,5*R*,8*R*)-5-Benzyl-8,11,11-trimethyl-3-oxa-6-azatricyclo-[6.2.1.0^{2,7}]undec-6-en-4-one (19e): 295 mg (yield 71%); [α]²²_D +227.2 (c = 1.94, CHCl₃); R_f 0.49 (hexanes/EtOAc = 4/1); mp 122–124 °C; IR (NaCl, CHCl₃) 2961 (m), 1747 (s), 1692 (m) cm⁻¹; ¹H NMR δ 7.42–7.20 (m, 5H), 4.43 (s, 1H), 3.96–3.92 (m, 1H), 3.54–3.49 (dd, J = 14.0, 5.2 Hz, 1H), 3.32–3.26 (dd, J = 14.0, 7.6 Hz, 1H), 2.24–2.22 (d, J = 4.8 Hz, 1H), 2.10–2.03 (m, 1H), 1.78–1.70 (m, 1H), 1.59–1.52 (m, 1H), 1.35–1.28 (m, 1H), 1.04 (s, 3H), 0.95 (s, 3H), 0.71 (s, 3H); ¹³C NMR δ 182.7, 170.7, 138.8, 130.0, 128.1, 126.3, 79.6, 62.5, 52.5, 49.1, 47.4, 37.4, 29.4, 25.3, 20.0, 19.6, 9.9; MS m/z 297 (M⁺, 83.6), 269 (27.2), 253 (21.5), 225 (13.4), 206 (27.8), 178 (46.0), 162 (33.7), 148 (19.0), 105 (16.7), 91 (100.0), 69 (35.4), 55 (16.3); HRMS m/e calcd for C₁₉H₂₃-NO₂ M⁺ 297.1732, found M⁺ 297.1729.

(1*S*,2*R*,5*S*,8*R*)-5-Allyl-5,8,11,11-tetramethyl-3-oxa-6-azatricyclo-[6.2.1.0^{2,7}]undec-6-en-4-one (19f): 274 mg (yield 75%); [α]²²_D +36.5 (c = 1.57, CHCl₃); R_f 0.42 (hexanes/EtOAc = 4/1); mp 85– 86 °C; ¹H NMR (200 MHz, CDCl₃) δ 5.82–5.69 (m, 1H), 5.18– 5.09 (m, 2H), 4.64 (s, 1H), 2.57–2.33 (m, 2H), 2.21 (d, J = 2.6, 1H), 2.12–2.00 (m, 1H), 1.85–1.50 (m, 2H), 1.62 (s, 3H), 1.42– 1.32 (m, 1H), 1.05 (s, 3H), 0.96 (s, 3H), 0.78 (s, 3H); ¹³C NMR δ 183.5, 171.8, 131.8, 120.0, 79.7, 63.5, 56.8, 52.3, 49.1, 42.8, 29.5, 25.2, 20.0, 19.7, 17.3, 10.0; MS m/z 261 (M⁺, 1.6), 217 (51.9), 202 (59.3), 192 (42.8), 189 (42.7), 176 (33.4), 148 (93.4), 124 (38.6), 107 (23.9), 91 (45.5), 69 (66.7), 53 (37.5). 41 (100); HRMS m/e calcd for C₁₆H₂₃NO₂ 261.1729, found 261.1728. Anal. Calcd for C₁₆H₂₃NO₂: C, 73.53; H, 8.87; N, 5.36. Found. C, 73.60; H, 8.88; N, 5.33.

General Procedure for Hydrolysis of α, α -Dialkylated Tricyclic Iminolactones. To a solution of 18 or 19 (0.4 mmol) in 10 mL of MeOH/acetone (v/v = 1:1) was added 2 N NaOH (0.5 mL, 1 mmol), and the mixture was stirred for 2 h at room temperature. The solution was diluted with 10 mL of water, cooled to 0 °C, and then acidified with 2 N HCl to pH 3–4. The acidic solution was extracted with ethyl acetate (3 × 15 mL). The organic layer was dried over anhydrous MgSO₄ and filtered. After removal of the solvent, the residue was transferred to a sealed tube and 6 N HCl (2 mL) was added. This mixture was heated at 92 °C for 3 h and then extracted with ethyl acetate (3 \times 5 mL). The aqueous layer was evaporated to dryness under reduced pressure, and the residue was dissolved in anhydrous ethanol (2 mL) followed by the addition of of propylene oxide (1.5 mL). The mixture was stirred at room temperature for 1 h (or heated at reflux for 30 min). After removal of the ethanol, the resulting white solid precipitate was washed with acetone (2 mL), collected by filtration, washed successively with acetone (3 \times 2 mL), and air-dried to afford the desired α,α -dialkylated- α -amino acid.

(S)-2-Methylphenylalanine: 63 mg; yield 88%; mp 278–280 °C; $[\alpha]^{22}_{D}$ –21.6 (c = 1.04, H₂O) [lit.²⁴ $[\alpha]^{20}_{D}$ –22 (c = 1 in H₂O)]; ¹H NMR (400 MHz, D₂O) δ 7.29–7.26 (m, 3H), 7.15 (d, J = 7.0 Hz, 2H), 3.19 (d, J = 14.2 Hz, 1H), 2.87 (d, J = 14.2 Hz, 1H), 1.44 (s, 3H).

(*R*)-2-Methylphenylalanine: 62 mg; yield 86%; mp 277–279 °C; $[\alpha]^{22}_{D}$ +21.5 (*c* = 1.04, H₂O); ¹H NMR (400 MHz, D₂O) δ 7.30–7.26 (m, 3H), 7.16 (d, *J* = 6.2 Hz, 2H), 3.20 (d, *J* = 14.2 Hz, 1H), 2.88 (d, *J* = 14.2 Hz, 1H), 1.44 (s, 3H).

(*S*)-2-Propenyl-phenylalanine: 67 mg; yield 82%; mp > 300 °C; $[\alpha]^{22}_{D}$ +26.8 (c = 1.11, H₂O) [lit.²⁵ $[\alpha]^{20}_{D}$ +27.3 (c = 1 in H₂O)]; ¹H NMR (400 MHz, D₂O) δ 7.29–7.25 (m, 3H), 7.17 (d, J = 7.5 Hz, 2H), 5.75–5.65 (m, 1H), 5.23 (s, 1H), 5.20–5.18 (d, J = 6.4 Hz, 1H), 3.26 (d, J = 14.8 Hz, 1H), 2.93 (d, J = 13.7 Hz, 1H), 2.76–2.73 (dd, J = 14.6, 6.4 Hz, 1H), 2.45–2.39 (dd, J = 14.6, 8.8 Hz, 1H).

(*R*)-2-Propenylphenylalanine: 71 mg; yield 87%; mp > 300 °C; $[\alpha]^{22}_{D} - 27.0$ (c = 1.10, H₂O); ¹H NMR (400 MHz, D₂O) δ 7.30–7.26 (m, 3H), 7.17 (d, J = 7.4 Hz, 2H), 5.68–5.62 (m, 1H), 5.21(s, 1H), 5.17 (d, J = 6.3 Hz, 1H), 3.24 (d, J = 14.8 Hz, 1H), 2.91 (d, J = 14.2 Hz, 1H), 2.74–2.69 (dd, J = 14.6, 6.4 Hz, 1H), 2.43–2.37 (dd, J = 14.6, 8.8 Hz, 1H).

(S)-2-Amino-2-methyl-4-pentenoic acid: 46 mg; yield 90%; mp > 300 °C; $[\alpha]^{22}_{D}$ -28.0 (c = 0.88, H₂O) [lit.²⁶ $[\alpha]^{25}_{D}$ -28.5 (13 mg/dm³, H₂O, liq, 10 cm)]; ¹H NMR (400 MHz, D₂O) δ 5.65-5.58 (m, 1H), 5.16 (s, 1H), 5.13-5.12 (d, J = 4.1 Hz, 1H), 2.56-2.51 (dd, J = 14.4, 6.5 Hz, 1H), 2.35-2.30 (dd, J = 14.4, 8.4 Hz, 1H), 1.36 (s, 3H).

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Supporting Information Available: X-ray position parameters, full bond distances and angles, and ORTEP drawings for **18b**, **18d**, **18g**, and **19e** as well as tables of total energies and Cartesian coordinates for the minimized structures by AM1 and ab initio calculations are included. This material is available free of charge via the Internet at http://pubs.acs.org.

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